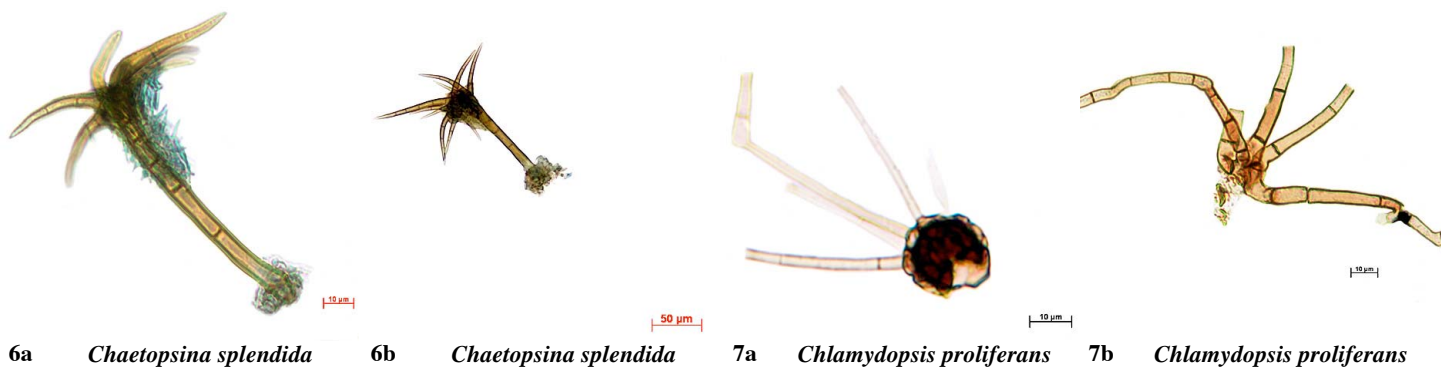
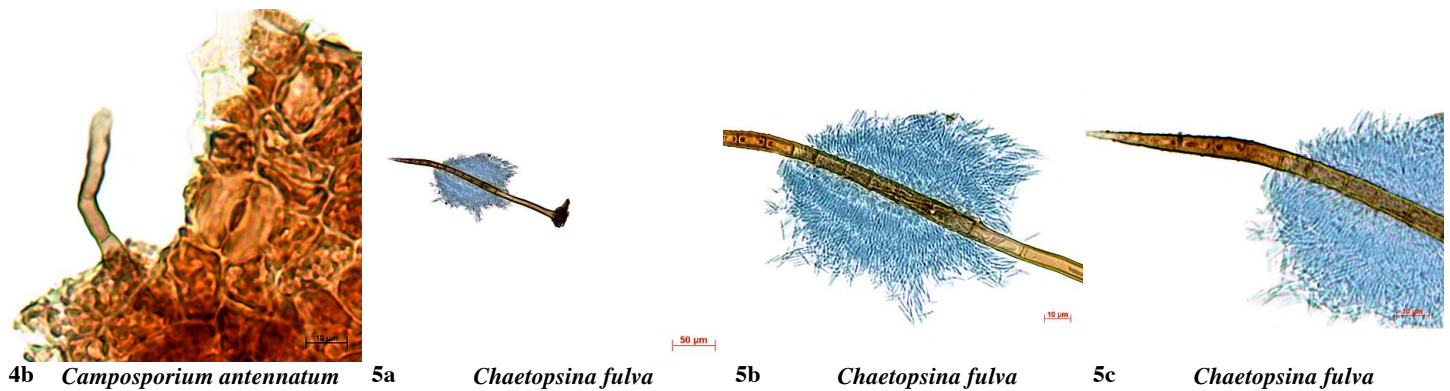
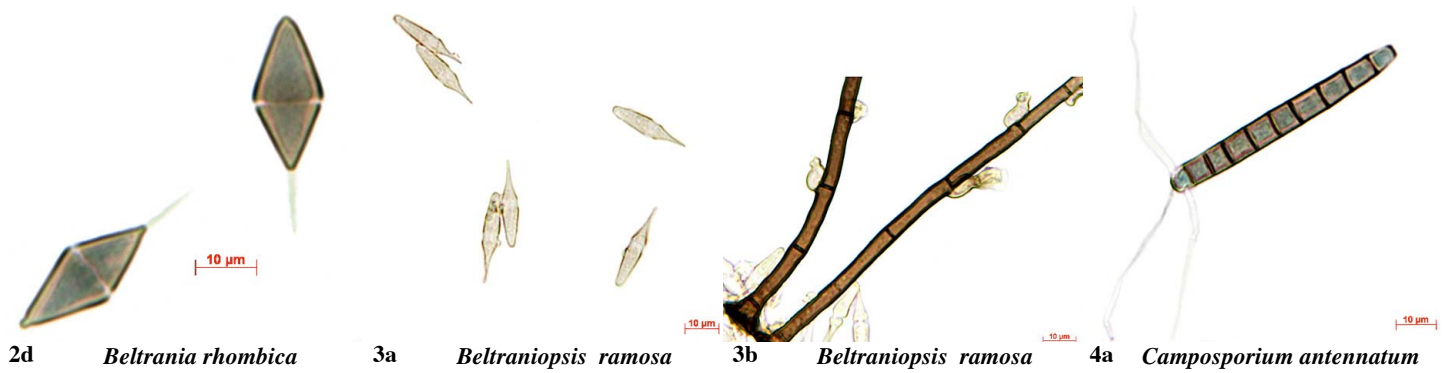
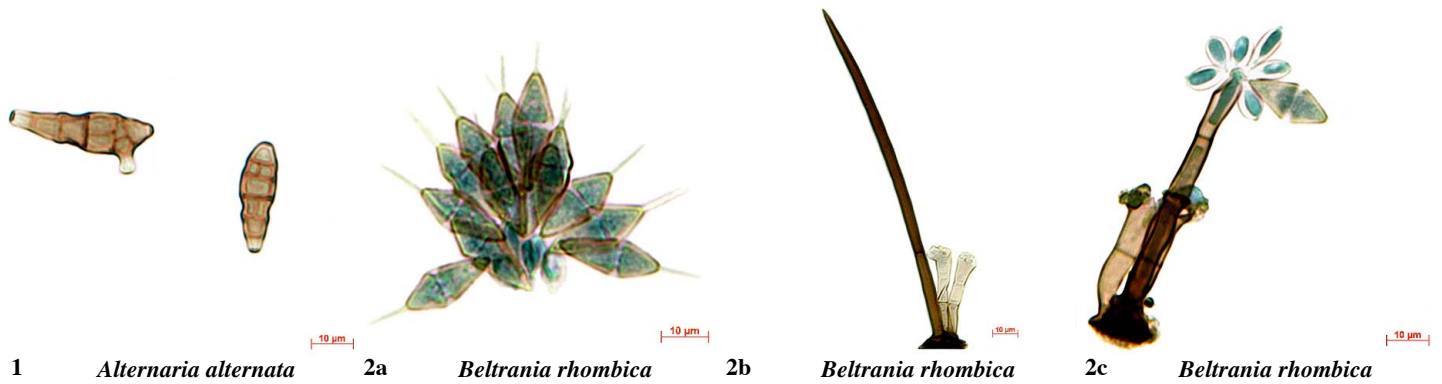


# Hyphomycetes decompositores de *Caesalpinia echinata* Lam. 1 no Estado de São Paulo, Brasil

Priscila da Silva e Rosely Ana Piccolo Grandi

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# Hyphomycetes decompositores de *Caesalpinia echinata* Lam. no Estado de São Paulo, Brasil

Priscila da Silva e Rosely Ana Piccolo Grandi

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8a *Circinotrichum olivaceum*

10 µm



8b *Circinotrichum olivaceum*

10 µm



9a *Composporium pellucidum*

10 µm



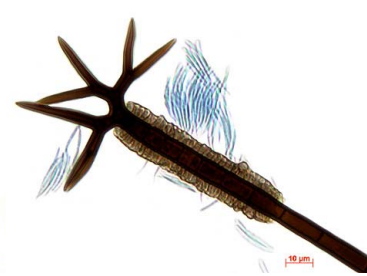
9b *Composporium pellucidum*

10 µm



10a *Cryptophiale udagawae*

50 µm



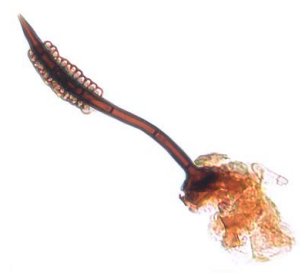
10b *Cryptophiale udagawae*

10 µm



11a *Cryptophiale minor*

10 µm



11b *Cryptophiale minor.*

10 µm



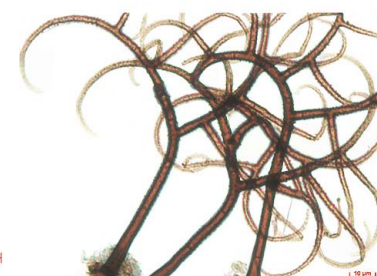
12a *Gyrothrix circinata*

50 µm



12b *Gyrothrix circinata*

10 µm



12c *Gyrothrix circinata*

10 µm



13a *Gonytrichum macrocladum*

50 µm



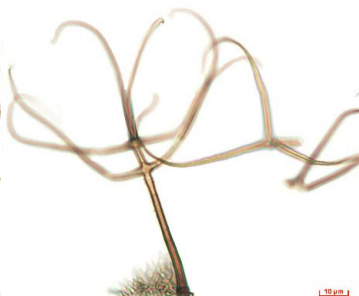
13b *Gonytrichum macrocladum*

10 µm



14a *Gyrothrix ramosa*

10 µm



14b *Gyrothrix ramosa*

10 µm



15 *Gyrothrix microsperma*

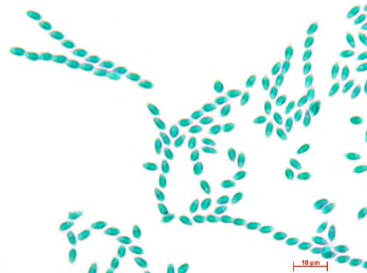
# Hyphomycetes decompositores de *Caesalpinia echinata* Lam. no Estado de São Paulo, Brasil

Priscila da Silva e Rosely Ana Piccolo Grandi

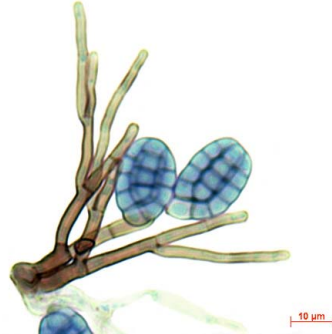
Fotos: Priscila da Silva e Rosely Ana Piccolo Grandi. Agradecimentos: FAPESP – Fundação de Amparo a Pesquisas do Estado de São Paulo, SP. Processo no 05/51732-5 e Dra. Marina Capelari  
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16a *Mariannaea elegans*



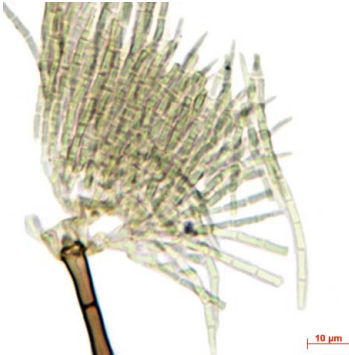
16b *Mariannaea elegans*



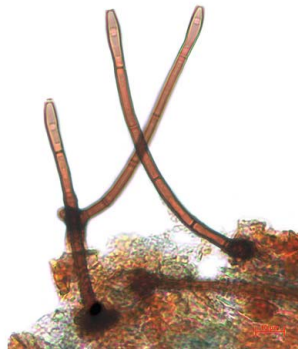
17 *Pseudodictyosporium wauense*



18a *Speiropsis scopiformis*



18b *Speiropsis scopiformis*



19a *Sporidesium flagelliforme*



19b *Sporidesium flagelliforme*



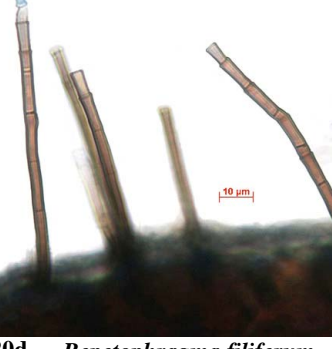
20a *Repetophragma filiferum*



20b *Repetophragma filiferum*



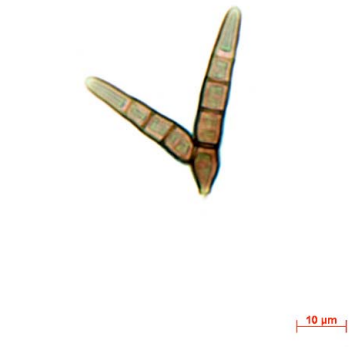
20c *Repetophragma filiferum*



20d *Repetophragma filiferum*



21a *Triposporium deviatum*



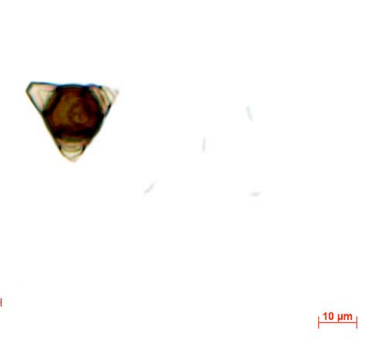
21b *Triposporium deviatum*



22a *Uberispora heteroseptata*



22b *Uberispora heteroseptata*

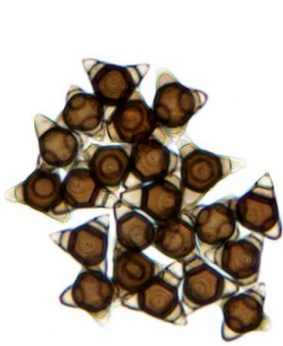


22c *Uberispora heteroseptata*

# Hyphomycetes decompositores de *Caesalpinia echinata* Lam. no Estado de São Paulo, Brasil

Priscila da Silva e Rosely Ana Piccolo Grandi

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22d *Uberispora heteroseptata*



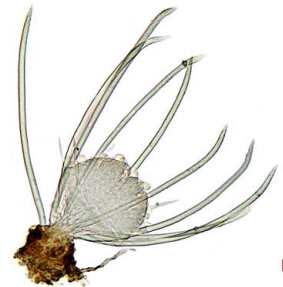
23a *Vermiculariopsiella immersa*



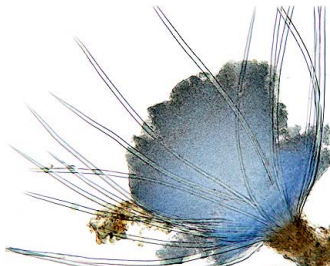
23b *Vermiculariopsiella immersa*



23c *Vermiculariopsiella immersa*



24a *Volutella minima*



24b *Volutella minima*



25a *Wiesneriomyces laurinus*



25b *Wiesneriomyces laurinus*



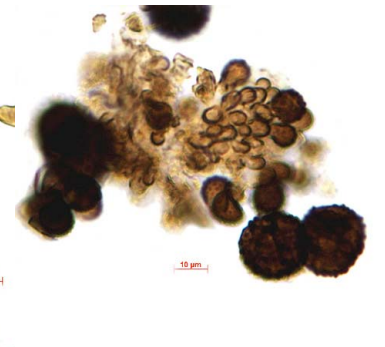
26 *Periconia minutissima*



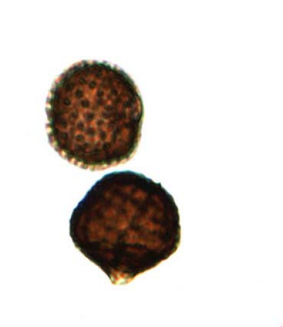
27a *Pithomyces maydicus*



27b *Pithomyces maydicus*



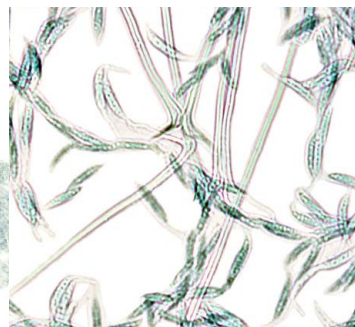
28a *Epicoccum nigrum*



28b *Epicoccum nigrum*



29a *Thozetella gigantea*



29b *Thozetella gigantea*



29c *Thozetella gigantea*

# Hyphomycetes decompositores de *Caesalpinia echinata* Lam. no Estado de São Paulo, Brasil

5

Priscila da Silva e Rosely Ana Piccolo Grandi

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1. <i>Alternaria alternata</i> (Fr.) Keissl - conidia muriform	16a. <i>Mariannaea elegans</i> (Corda) Samson – apical part of conidiophore with conidiogenous cells and conidia
2a. <i>Beltrania rhombica</i> Penz - conidia in group	16b. <i>Mariannaea elegans</i> (Corda) Samson – conidia catenulate
2b. <i>Beltrania rhombica</i> Penz - setae, conidiophores and conidiogenous cells terminal	17a. <i>Pseudodictyosporium wauense</i> Matsush – conidiophore, conidiogenous cells terminal and conidia setiform
2c. <i>Beltrania rhombica</i> Penz - conidiophores, conidiogenous cells terminal, separating cells ellipsoidal obovoid and conidium	18a. <i>Speiropsis scopiformis</i> Kuthub. & Nawawi – conidiophore setiform, conidiogenous cells terminal and conidia catenulate
2d. <i>Beltrania rhombica</i> Penz - conidia	18b. <i>Speiropsis scopiformis</i> Kuthub. & Nawawi – apical part of conidiophore with conidiogenous cells terminal and conidia catenulate
3a. <i>Beltraniopsis ramosa</i> R.F. Castañeda - conidia	19a. <i>Sporidesmium flagelliforme</i> Matsush – conidiophore and conidiogenous cells terminal
3b. <i>Beltraniopsis ramosa</i> R.F. Castañeda – conidiophores and conidiogenous cells doliform	19b. <i>Sporidesmium flagelliforme</i> Matsush - conidium
4a. <i>Camposporium antennatum</i> Harkn - conidium	20a. <i>Repetophragma filiferum</i> (Piroz) R. F. Castañeda - conidium
4b. <i>Camposporium antennatum</i> Harkn - conidiophore	20b. <i>Repetophragma filiferum</i> (Piroz) R. F. Castañeda – conidiogenous cells percurrent and conidium
5a. <i>Chaetopsina fulva</i> Rambelli – conidiophore setiform with conidia in the median region	20c. <i>Repetophragma filiferum</i> (Piroz) R. F. Castañeda - conidia
5b. <i>Chaetopsina fulva</i> Rambelli – conidiogenous region with conidia	20d. <i>Repetophragma filiferum</i> (Piroz) R. F. Castañeda - conidiophores
5c. <i>Chaetopsina fulva</i> Rambelli – apical part of conidiophore with conidia	21a. <i>Triposporium deviatum</i> (Subram.) R. F. Castañeda – conidiophores, conidiogenous cell terminal and conidium
6a. <i>Chaetopsina splendida</i> B. Sutton & Hodges – conidiophore setiform and conidia	21b. <i>Triposporium deviatum</i> (Subram.) R. F. Castañeda - conidium
6b. <i>Chaetopsina splendida</i> B. Sutton & Hodges – conidiophore setiform	22a. <i>Uberispora heteroseptata</i> R. F. Castañeda, Guarro & Cano – conidiophore, conidiogenous cell terminal and conidia
7a. <i>Chlamydopsis proliferans</i> Hol –Jech & R. F. Castañeda - conidia	22b. <i>Uberispora heteroseptata</i> R. F. Castañeda, Guarro & Cano – conidiogenous cell percurrent with conidium
7b. <i>Chlamydopsis proliferans</i> Hol –Jech & R. F. Castañeda – conidiophores with conidiogenous cells terminal	22c. <i>Uberispora heteroseptata</i> R. F. Castañeda, Guarro & Cano – conidium and phialoconidia
8a. <i>Circinotrichum olivaceum</i> (Speg.) Piroz - seta	22d. <i>Uberispora heteroseptata</i> R. F. Castañeda, Guarro & Cano - conidia
8b. <i>Circinotrichum olivaceum</i> (Speg.) Piroz – setae in group	23a. <i>Vermiculariopsiella immersa</i> (Desm.) Bender – conidiomats with setae and conidia
9a. <i>Camposporium pellucidum</i> (Grove) S. Hughes - conidiophore	23b. <i>Vermiculariopsiella immersa</i> (Desm.) Bender – conidiomats with setae and conidia
9b. <i>Camposporium pellucidum</i> (Grove) S. Hughes - conidium	23c. <i>Vermiculariopsiella immersa</i> (Desm.) Bender – conidiomats with setae and conidia
10a. <i>Cryptophiale udagawae</i> Piroz & Ichinoe – conidiophore setiform	24a. <i>Volutella minima</i> Höhn – conidiomats with setae and conidia
10b. <i>Cryptophiale udagawae</i> Piroz & Ichinoe – apical part of conidiophore with conidiogenous conidia	24b. <i>Volutella minima</i> Höhn – conidiomats with setae and conidia
11a. <i>Cryptophiale minor</i> M.L. Farr – conidiophores setiform and conidia	25a. <i>Wiesneriomyces laurinus</i> (Tassi) P. M. Kirk – conidiomats with setae and conidia
11b. <i>Cryptophiale minor</i> M.L. Farr – conidiophore setiform and conidiogenous region	25b. <i>Wiesneriomyces laurinus</i> (Tassi) P. M. Kirk – conidia catenulate
12a. <i>Gyothyrix circinata</i> (Berk. & M. A. Curtis) S. Hughes – setae in group	26. <i>Periconia minutissima</i> Corda – conidiophore, conidiogenous cells terminal and conidia
12b. <i>Gyothyrix circinata</i> (Berk. & M. A. Curtis) S. Hughes – setae and conidia	27a. <i>Pithomyces maydicus</i> (Sacc.) M. B. Ellis – conidia on detritus
12c. <i>Gyothyrix circinata</i> (Berk. & M. A. Curtis) S. Hughes - setae	27b. <i>Pithomyces maydicus</i> (Sacc.) M. B. Ellis - conidia
13a. <i>Gonytrichum macrocladum</i> (Sacc.) S. Hughes – conidiophores setiform	28a. <i>Epicoccum nigrum</i> Link - conidia
13b. <i>Gonytrichum macrocladum</i> (Sacc.) S. Hughes – conidiophore setiform, conidiogenous cells and conidia	28b. <i>Epicoccum nigrum</i> Link - conidia
14a. <i>Gyothyrix ramosa</i> Zucconi & Onofri – setae and conidiogenous cells at the base	29a. <i>Thozetella gigantea</i> B. C. Paulus, Gadek & K. D. Hyde – synnema without synchronous proliferation and conidia in mucilaginous mass.
14b. <i>Gyothyrix ramosa</i> Zucconi & Onofri – setae and conidiogenous cells at the base	29b. <i>Thozetella gigantea</i> B. C. Paulus, Gadek & K. D. Hyde – lunate conidia and microawns in L-shaped
15. <i>Gyothyrix microsperma</i> (Höhn.) Piroz -setae	29c. <i>Thozetella gigantea</i> B. C. Paulus, Gadek & K. D. Hyde – L-shaped microawns